

# Potential Alcohol Production from Beta Vulgaris Genotypes as Affected by Nitrogen Level and Water Stress

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## INTRODUCTION

Present and future world shortages and increasing petroleum costs have stimulated the search for alternate renewable and nonrenewable energy sources. Sugarbeets (*Beta vulgaris* L.) and sugarcane (*Saccharum spp.*) have high potential as a feedstock for conversion to alcohol as a practical renewable energy source. Sugarbeets have many desirable characteristics such as: storage of 40 to 50% of their dry matter as fermentable sugars (6, 7, 15); a small nitrogen (N) requirement per unit of sugar produced (2, 5, 7); a range of related *Beta vulgaris* genotypes which may be used to increase yield potential (7, 8); use of by-products as a cattle feed or conversion to methane (6, 7); and wide adaptation within the U.S. (19). They can also be stored up to 6 months in cool areas, all of which make them a primary feedstock source for alcohol production.

In the development of new sugarbeet cultivars for sucrose production, high sucrose concentration with low levels of impurities including nonsucrose sugars in the harvested root has been emphasized to maximize extractable sucrose. Fodder beets, which in contrast have higher root yields, high impurities, and low sugar concentration, have been grown extensively in Europe as cattle feed. Many of the disadvantages to fodder beets as a refined sucrose source are not important if they are to be used for alcohol production. Although low in sugar concentration, the high root yielding fodder beet may have the potential to

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yield more fermentable sugars per hectare than can be obtained from commercial sugarbeet varieties.

Soil tests have been developed to assess the residual N and provide a basis for calculating N fertilizer application levels needed to maximize the product of root growth and extractable sucrose concentration of commercial sugarbeet hybrids (2, 5, 9, 10, 11). However, little is known concerning the N requirements of other genotypes for situations where total fermentable sugar production is of primary importance.

The sugarbeet water requirements have been studied extensively and reviewed by Jensen and Erie (12). Recent studies show that irrigation water level can be reduced below present day practices during the growing season with very little effect on sucrose yield (3, 4, 18, 22). The areas of the western U.S. where many beets could be grown for alcohol production may have limited irrigation water.

The objective of this study was to evaluate the fermentable sugar yield potential of several genetically diverse *Beta vulgaris* genotypes (sugarbeet and fodder beet types) grown under moderate and high N levels and normal and mid-to late-season soil water deficit and plant water stress and to determine their N and water requirements.

#### MATERIALS AND METHODS

An irrigated field experiment was conducted on Portneuf silt loam soil (Durixerollic Calcorthids; coarse-silty, mixed, mesic) near Twin Falls, Idaho in 1980. This soil has a weakly cemented hardpan at the 50- to 60-cm depth that has little effect on water movement when saturated but may restrict root penetration. The area used was cropped to barley (*Hordeum vulgare* L.) the previous year and was deficient in N (2) and phosphorus (P) (21).

Two adjacent experimental areas (irrigation treatments), each involving six replications of a split-plot design with two N rates as main plots and eight *Beta vulgaris* genotypes as subplot plots, were used. The N treatments were broadcast preplant as ammonium nitrate at 196 and 392 kg N/ha on plots 17.9 m wide by 7.6 m long. Phosphorus was applied uniformly at 56 kg P/ha. The fertilizers were incorporated with the upper 10

cm of soil as the seedbed was prepared.

The N fertilizer rates used were established by the use of the sugarbeet N requirement equation developed by Carter et al. (2, 5).

$$N_f = \frac{Y_E (5.5 \pm 0.5) - (\alpha_{nN} + \alpha_{mN} - 5 R_s)}{E_f} \quad [1]$$

where  $N_f$  is the N fertilizer/ha needed,  $Y_E$  is the expected maximum yield,  $5.5 \pm 0.5$  kg of N is the N required to produce a metric ton of crowned fresh roots (root minus crown),  $\alpha_{nN}$  is the available  $NO_3^-N$  in the soil depth sampled,  $\alpha_{mN}$  is the available mineralizable N in the soil depth sampled,  $R_s$  is the straw in metric tons/ha, and  $E_f$  is the efficiency of applied N fertilizer.

Eight *Beta vulgaris* genotypes (Table 1) were planted in four row plots on 8 April 1980. All genotypes were planted in 56-cm rows that had previously been marked and treated with aldicarb at 2.24 kg of active ingredient per hectare to control insects. The genotypes were thinned to a 23- to 30-cm within-row spacing in early June.

Two irrigation levels,  $M_1$  and  $M_3$  (3), were used. The irrigation times and amounts are summarized in Figure 1 for the following treatments:

$M_1$  - Adequate irrigation based on previous experiments. Irrigation dates were based on estimated soil moisture depletion (13) and irrigation durations depended on the amount to be applied.

$M_3$  - No irrigation was applied after the soil profile was filled with water on 1 August. Irrigations were the same as  $M_1$  before 1 August.

A light irrigation was applied to both irrigation levels about 10 days before harvest. Every other row irrigation was used throughout the season. The net amount of water applied was determined using an equation previously reported (3).

The soil water content in the 0- to 30-cm depth was determined gravimetrically at various times between 30 July to 6 October. One access tube located within the center rows of each of the genotypes on two replications (196 kg N/ha) near the upper and lower part of each irrigation treatment and a calibrated neutron probe were used to measure the soil moisture in

IRRIGATIONS AND RAINFALL<sup>1</sup>

Treatment to 30 June	July	Aug.	Sept.	Oct.	Total
Rainfall	1 70 70	1 70 70	1 70 70	1 70 70	1 79 79
M <sub>1</sub>	211				46 <sup>2</sup>
M <sub>3</sub>	211	70 70	180 70		890 680

<sup>1</sup> Arrows above quantity of water refers to application date. \* After 1 August.

Figure 1. Irrigation water applied and rainfall. the 30- to 230-cm depth at the same time intervals as of those taken for the 0- to 30-cm depth between 30 July to 6 October. Petiole samples consisting of 25 of the youngest fully-mature leaves were selected at random from each plot of the M<sub>1</sub> irrigation treatment on 21 August. The petioles were cut into 0.5-cm sections, dried at 65°C, ground to pass through a 40-mesh sieve, subsampled, and analyzed for NO<sub>3</sub>-N, using a nitrate specific ion electrode (17).

Root and top samples were harvested manually from three uniform 3-m row sections from the center rows of each plot between 14 to 21 October. Root samples that included the crown were brushed free of soil, weighed, and triplicate root samples consisting of 10 to 12 roots were taken for total sugars, sucrose, purity, dry matter, and N analysis. A representative top sample was taken from a composite of the three row sections for dry matter and N analysis. All fresh tissue was weighed before and after drying. Extra root samples were taken from the genotypes AH10 and GWD2 to determine the yield of crowded roots (root minus crown).

Total sugars, reducing sugars, and sucrose were determined by the Sugarbeet Research Laboratory at Logan, Utah, using the following methods: sucrose concentration was determined by the Sach - le Docte cold digestion procedure as outlined by McGinnis (16), reducing sugars (glucose and fructose) were quantified colorimetrically in the filtrate obtained for sucrose analysis using dinitrosalicylic acid reagent. Total fermentable sugars are the sum of sucrose and reducing sugars. Dry matter in the beet roots was determined by drying brei samples collected at the

<sup>2</sup> 2n sugarbeet x 2n fodder beet.  
+ X = highly susceptible, XX = partial resistance, XXX = highly resistant.

Name	Seed	Frostmelt	Cultivar	Top†	German	Description
AH10	U.S.	XX	mono-	Commercial hybrid, Great Western Sugar Co.		
GWD2	U.S.	XXX	mono-	Commercial hybrid, Amalgamated Sugar Co.		
LHY-1	U.S.	XXX	multi-	Logan high sucrose conc., exp., hybrid.		
LHS-1	U.S.	XXX	multi-	Logan high yield exp., hybrid.		
Monososa	Nettelerlands	X	mono-	Diploid high yield hybrid beet		
Monoblancc	Nettelerlands	X	mono-	Diploid high yield hybrid beet		
Pajibjergr Kortsoe	Denmark	X	multi-	Triplloid high yield hybrid beet		
Rota	Germany	X	multi-	Diploid open pollinated beet		

Table 1. Description of Beta vulgaris genotypes used in this experiment.

time of sugar and sucrose analyses.

Samples from the tops and roots (root plus crown) were dried at 65°C and dry weight determined. The dried samples were ground to pass a 40-mesh sieve and the total N was determined by the semimicro-Kjeldahl procedure modified to include nitrate (1). Nitrogen uptake was calculated by assuming that the N concentration was the same in both the fibrous and storage roots and that the weight of the unharvested fibrous roots was equal to 25% of the total harvested storage root weight (14).

#### RESULTS AND DISCUSSION

The growth patterns of the various genotypes were very similar throughout the growing season. The main differences were in early and late top growth and the position of the storage roots in relation to the soil surface. From field observations during mid-June to mid-July, the earliest and greatest top growth was observed for Pajbjerg Korsroe, Monoblanc, and Rota; followed by GWD2 and Monorosa; and smallest by the AH10, LHY-1, and LHS-1. The genotypes established complete cover of the soils surface by the leaf canopy in the same order during early to mid-July. Plants at the higher N level had a larger leaf canopy at all growth stages on all genotypes as compared with the lower N treatment. The fodder beets (Pajbjerg Korsroe and Rota) started to turn yellow in mid-August on the lower N treatment indicating a N deficit. This was particularly noticeable on Rota where only the younger leaves were green at both N levels by the end of the season. Leaf canopy was reduced on both N levels and all genotypes by mid- to late-season moisture stress as compared with the normal irrigation. In sugarbeet types, only the crown of the storage root is above the soil surface. The genotypes Monoblanc, Pajbjerg Korsroe, and Rota had 40 to 60% of the storage root above the soil surface at harvest. This could have adverse effects on harvesting the whole storage root for alcohol production using present day farm equipment.

The genotypes varied widely in their root yield, and sucrose and total sugar concentration at both N levels (Table 2). The fodder beets (Pajbjerg Korsroe and Rota) gave significantly higher root yields, lower sucrose and total sugar concentrations as compared with the sugarbeets (GWD2, AH10, LHY-1, and LHS-

Table 2. Interaction means for each genotype at each N level and N level means for root yield, sucrose (% and yield), total sugar (% and yield), and potential alcohol yield; mc = metric tons, kl = kiloliters.

	Genotype	Root Yield mc/ha	Root Yield 196†	Root Yield 392†	Sucrose 196	Sucrose 392	Total Sugars 196	Total Sugars 392	Alcohol† 196	Alcohol† 392			
GWD2		—	—	—	%	%	— mt/ha —	— mt/ha —	— kl/ha —	— kl/ha —			
AH10		74.3	* 80.3	18.2	* 16.9	14.5	14.4	18.4	* 17.1	14.6	14.5	8.72	8.67
LHY-1		83.1	86.1	18.2	* 16.6	13.5	13.4	18.3	* 16.8	13.6	13.5	8.13	8.05
LHS-1		67.7	* 74.5	18.5	* 17.2	15.4	14.8	18.6	* 17.4	15.5	15.0	9.23	8.92
Monorosa		88.4	* 100.9	20.3	* 18.6	13.8	13.8	20.5	* 18.8	13.9	14.0	8.27	8.33
Monoblanc		102.3	* 110.5	15.2	* 14.1	13.5	14.2	15.4	* 14.2	13.6	* 14.4	8.10	8.56
Pajbjerg Korsroe		121.5	* 134.9	110.5	* 12.7	14.7	14.0	14.5	* 12.9	14.8	14.2	8.84	8.49
Rota		116.4	* 138.2	10.2	* 9.1	11.9	12.5	10.5	* 9.4	12.2	* 13.0	7.60	9.10
LSD (0.05)		3.8	3.8	0.5	0.5	0.5	0.5	0.8	0.5	0.7	0.7	0.45	0.45
N effect		*	*	NS	NS	NS	*	*	*	*	*	*	*
N interaction		91.7	* 101.3	15.8	* 14.5	13.9	14.0	16.0	* 14.7	14.1	14.2	8.40	8.49
N significant interaction at p = 0.05, between N level means = significant difference between means at p = 0.05.													
+ 1.66 kg sugar/litter alcohol (14 lbs/gal).													
kg N/ha.													

\* Significant interaction at p = 0.05, between N level means = significant difference between means at p = 0.05.

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- 1). The fodder  $\times$  sugarbeet hybrids (Monorosa and Monoblanc) were intermediate in root yields, sucrose and total sugar concentrations. This negative correlation between root yield and sucrose or total sugar concentrations resulted in similar yields of total sugar and potential alcohol production at both N levels. The highest potential alcohol production came from the experimental sugarbeet hybrid LHY-1, followed by the fodder beet hybrid Pajbjerg Korsroe. The lowest potential alcohol production was the highest root yielding genotype Rota, an open-pollinated fodder beet.

The fodder beets had the highest reducing sugar concentration on a dry (Table 3) and fresh root basis, but in no case did they exceed 0.5% on a fresh root basis. Total sugar concentration on a dry weight basis and root dry matter concentration were highest on the sugarbeets and lowest on the higher root yielding fodder beets with intermediate levels on the fodder  $\times$  sugarbeet hybrids. Total dry matter production was similar between the two divergent types (sugarbeets and fodder beet types) except for the nonhybrid fodder beet, Rota, which was significantly less. The proportions of the total dry matter in the tops and roots indicated that fodder beets partitioned less photosynthate to the tops and more to the roots than sugarbeets. These data also indicated that the fodder beets partitioned the photosynthetic energy in the roots differently than sugarbeets with less going for sugar storage.

The parameters that make up total sugar yield for the various genotypes were compared in a regression analysis at both N and water levels in Table 4. There was an inverse linear relationship between root yield and dry matter or total sugar concentration in the roots at both irrigation levels within each N level. The total sugar concentration in the fresh roots was primarily dependent upon the dry matter concentration within the roots with a lesser but still important total sugar concentration within the dry matter. Increased N level reduced the total sugar concentration in the fresh roots by reducing the percent dry matter and the total sugar concentration of the dry matter. There was no effect of irrigation level on the total sugar concentrations in the fresh and dry roots or the dry mat-

	N effect means										N effect means										
	Interaction					NS					Interaction					NS					
	1.07	*	1.30	74.6	*	72.8	21.4	*	20.2	4.4	*	6.2	18.9	*	19.6	23.3	*	25.8			
LSD (0.05)	0.11	0.11	0.11	2.4	0.5	0.5	0.3	0.3	1.1	1.1	1.2	1.2	1.2	1.2	1.2	1.2	1.2	1.2	1.2	1.2	
Rota	2.22	*	2.81	73.0	*	70.3	14.4	*	13.4	3.0	*	4.0	16.7	*	18.5	19.7	*	22.6	*	26.1	
Pajbjerg Korsroe	2.44	*	1.31	74.9	73.0	19.3	16.2	16.0	3.5	4.5	19.7	*	21.6	23.2	*	25.6	*	25.7	*	26.0	
Monoblanca	0.85	*	0.83	75.1	73.4	20.5	*	19.4	4.2	6.2	19.8	19.8	19.6	22.2	*	26.0	*	26.9	*	26.5	
Monorosa	0.67	*	0.76	75.2	75.2	24.8	*	23.3	27.3	4.9	*	6.9	18.5	19.0	23.4	*	26.0	*	26.5	*	26.9
LHS-1	0.61	*	0.67	75.2	72.1	24.7	*	23.3	5.7	6.8	20.6	20.6	25.4	24.1	*	26.5	*	26.9	*	26.0	
LHY-1	0.60	0.70	0.68	75.7	74.7	24.3	*	22.8	5.0	*	7.4	19.3	19.3	24.3	*	26.9	*	26.5	*	26.0	
GMD2	0.57	*	0.68	75.7	74.7	24.3	*	22.8	5.0	*	7.4	19.3	19.3	24.3	*	26.9	*	26.5	*	26.0	
Beta vulgaris	196+	392+	196	392	196	392	196	392	196	392	196	392	196	392	196	392	196	392	196	392	
Reducing Sugars	Total Sugars	Dry Matter	Root	Dry Matter	Tops	Dry Matter	Roots	Dry Matter	Tops	Dry Matter	Roots	Dry Matter	Roots	Dry Matter	Roots	Dry Matter	Roots	Dry Matter	Roots	Dry Matter	
Sugars as a percentage of dry matter, roots and tops, and total plant dry matter.	Yielded DM = dry matter.																				

Table 3. Interaction means for each genotype at each N level and N level means for reducing sugar and dry matter.

\* Significant interaction at  $p = 0.05$ , between N levels means = significant difference between means at  $p = 0.05$ .

+ kg N/ha.

ter concentration of the various genotypes. Therefore, total sugar yield was primarily dependent upon the dry matter yield at the different irrigation and N levels.

Table 4. Effect of root yield on percent dry matter and percent total sugars, percent root dry matter on percent total sugars, and dry matter yield of roots on total sugar yield as affected by N fertilizer level, and mid-to-late-season moisture stress.

N kg/ha	Fertilizer	$M_1$		$M_3$		Regression equation $r$	Regression equation $r$
		Root yield (yd)	Root yield (yd) on % Dry Matter†	Root yield (yd)	Root yield (yd) on % Total Sugar‡		
196		$\hat{y} = 41.1 - 0.210$	0.92	$\hat{y} = 39.1 - 0.197$	0.91	0.91	
392		$\hat{y} = 36.3 - 0.154$	0.95	$\hat{y} = 36.2 - 0.163$	0.93	0.93	
196		$\hat{y} = 31.1 - 0.160$	0.91	$\hat{y} = 29.7 - 0.154$	0.90	0.90	
392		$\hat{y} = 27.8 - 0.123$	0.94	$\hat{y} = 26.6 - 0.123$	0.93	0.93	
196		$\hat{y} = -0.41 + 0.770$	DM 0.99	$\hat{y} = -0.65 + 0.773$	DM 0.98	0.98	
392		$\hat{y} = -1.12 + 0.794$	DM 0.98	$\hat{y} = -0.31 + 0.736$	DM 0.98	0.98	
196		$\hat{y} = -1.47 + 0.826$	YDM 0.96	$\hat{y} = 0.30 + 0.725$	YDM 0.89	0.89	
392		$\hat{y} = 1.20 + 0.676$	YDM 0.85	$\hat{y} = 0.69 + 0.683$	YDM 0.82	0.82	
$\dagger S_b$ (Common standard error of the slopes) = 0.011,		$\ddagger S_b$ = 0.009,					
$\ddagger S_b = 0.021$ ,		$\ddagger S_b = 0.055$ .					

Nearly all the measured factors were affected by increasing the N level on all genotypes (Tables 2, 3). High N levels increased root yield and reducing sugars but decreased the concentrations of sucrose, total sugar and dry matter. This resulted in little difference in total sugars and potential alcohol production between the two N levels except for the fodder beet types.

Genotype  $\times$  N interactions were largely due to the difference in N treatment effects on the fodder and sugarbeet types. The fodder beets (Pajbjerg Korsroe and Rota) showed a much greater root yield increase and a smaller decrease in total sugar concentration at the higher N level than sugarbeets. This resulted in significant increases in total sugar and potential alcohol production for the fodder beets at the high N level; whereas the sugarbeets showed no difference at the two N levels.

The genotypes produced more dry matter under the high N level with the tops generally showing the greatest increase. The fodder beets yielded significantly more root dry matter at the high N level; whereas the sugarbeets produced essentially the same root dry matter at the two N levels. There was very little difference between the sugarbeet hybrids for total dry matter production at the high N level. The increased dry matter production caused by the higher N levels was mainly partitioned to top growth in the sugarbeet types; whereas in the fodder beet types the extra dry matter and growth was equally divided between the tops and roots.

The N requirement for maximum fermentable sugars and potential alcohol production for the various genotypes, using data from the optimum irrigation level ( $M_1$ ), were evaluated in Table 5. Previous experiments (5) have shown, because of the linear decrease in sucrose concentration with increasing total available N ( $N_T$ ), that maximum sucrose yield was obtained at a  $N_T$  value that was slightly less than required for maximum root yield. The same criteria should apply for total sugar yield on the various genotypes. The large increase in root yields that occurred for most genotypes with increased N rate indicated that maximum sugar yields would probably occur somewhere between the two application rates of 196 and 392 kg N/ha.

The two N application levels were calculated using a measured  $\alpha_{N_n} + \alpha_{m} N_m$  value of 146 kg N/ha and the upper limit of 6 kg N/metric ton in Equation [1] as the optimum N rates for a yield of 57 metric tons (mt) of crowned roots (root minus crown) per hectare for the genotype AH10 if a N fertilizer efficiency ( $E_f$ ) of 100 and 50% was obtained for the 196 and 392 kg N/ha rates, respectively. From past experiments (2, 5), an average  $E_f$  of 65% has been determined and used. However, in this experiment  $E_f$  for AH10 was found to be 75% at both N rates. The maximum root yield for the crowned AH10 roots of 71 metric tons/ha was found to be considerably above that used to determine N application rates. If the measured root yield for 392 kg N/ha level is used to determine the N requirements per metric ton of roots, the required kg N/metric ton would be 4.8, 5.6, 6.2, and 7.6 for the 50, 65, 75, and 100%  $E_f$ , respectively. Past

experimental work has shown the  $5.5 \pm 0.5$  kg N/metric ton is the optimum N rate for maximum sucrose yield. However, the average N requirement using the actual root yield of the 196 and 392 kg N/ha fertilizer levels for AH10 (roots minus crown) at 75%  $E_f$  would be near 5.3 kg N/metric ton (Table 5).

The total N uptake for the various genotypes was quite similar and averaged 258, 274, and 260 kg N/ha on the 196 kg N/ha rate and 400, 429, and 411 kg N/ha on the 392 kg N/ha rate for the sugarbeet, fodder  $\times$  sugarbeet, and fodder beet types, respectively. Although total N uptakes were similar, a greater proportion of the N was found in the roots of the fodder  $\times$  sugarbeet and fodder beet types as compared with sugarbeet types (Table 5). The N uptake used to produce a metric ton of sugar was significantly greater at the 392 kg N/ha level than at the 192 kg N/ha level. However, there were no significant differences between genotypes in N uptake per metric ton of sugar produced except for Rota at the 192 kg N/ha level.

Similarity between the total N uptake by the various genotypes indicated that the N necessary for maximum total sugar yield would also be similar. This is further verified by the petiole  $\text{NO}_3^-$ -N in late August (Table 6). The critical low range for  $\text{NO}_3^-$ -N has been established at 1000 ppm (20) and experience in this area indicates that petiole  $\text{NO}_3^-$ -N should be near 1000 ppm by late August to maximize yields, sucrose concentration and purity of sugarbeets. In the present work, the petiole  $\text{NO}_3^-$ -N concentration in late August was found to be quite similar between the various genotypes at each N level. Based on past experience, the 196 kg N/ha rate was found too low in petiole  $\text{NO}_3^-$ -N and 392 kg N/ha rate in slight excess for maximum sucrose and total sugar yields. This would indicate that the same criteria used for determining N needs of sugarbeets may be applicable to fodder  $\times$  sugarbeet and fodder beet types.

If the sugarbeet N requirements are applicable to fodder  $\times$  sugarbeet and fodder beet types as they appear to be, the average kg N/metric ton value at 75% N fertilizer efficiency for the two fertilizer rates (196 and 392 kg N/ha) in Table 5 should be used in Equation [1] to determine the N needs to maximize sugar production for the various genotypes. The  $E_f$  value in

Genotype	196 kg N/ha		392 kg N/ha		196 kg N/ha		392 kg N/ha		196 kg N/ha		392 kg N/ha		
	Tops	Roots											
GWD2	4.1	6.1	3.5	5.0	4.3	3.3	38	67	62	3.0	4.8	16.5	27.5
GWD21	4.6	6.8	3.9	5.5	4.7	3.3	33	67	62	3.0	4.8	—	—
AH10	4.5	6.4	3.9	5.3	4.7	3.6	36	41	64	59	3.4	5.3	29.0
AH101	5.2	7.6	4.4	6.2	5.3	4.6	36	41	64	59	4.9	5.7	—
LHY-1	4.0	6.1	3.5	5.0	4.3	32	32	41	68	59	3.7	5.7	—
LHS-1	5.0	7.0	4.3	5.7	5.0	31	37	69	63	3.2	4.2	17.7	26.7
Monojosa	3.7	5.2	3.2	4.2	3.7	22	27	78	73	3.6	4.2	19.6	30.4
Monoblanco	3.2	4.7	2.7	3.8	3.3	27	31	73	69	2.5	3.6	17.2	27.8
Pajibjerego Kortsore	2.8	3.8	2.4	3.1	2.8	23	22	77	78	2.2	3.2	18.6	28.1
Roca	2.9	3.8	2.5	3.1	2.8	17	20	83	80	2.2	2.6	21.4	28.0

† Residual  $\text{NO}_3^-$ -N and mineralizable N = 146 kg N/ha. ‡ N fertilizer efficiency. § kg N/ha.

%

kg N/metric ton.

Equation [1] can be expected to range from 50 to 75% depending on management practices (2) and it was found to average 65% in previous studies. The main difficulty would be that fodder x sugarbeet and fodder beet types have not been grown in the U.S. for a sufficient number of years to establish the root yield potential for the individual climatic zones. Until the root yield potentials are established, it would be preferable to use the N test for sugarbeets where the root yield potentials are generally known. However, there may be yield benefits to a slightly higher  $N_T$  level for fodder beets.

Sugarbeet water requirements have been studied extensively (12) and the general consensus for irrigated areas is that early light irrigations are needed to assure seed germination to establish and maintain a good stand with vigorous early growth. Soil water for the remainder of the season should be maintained at a favorable level to allow sufficient top growth and maintain leaf turgidity so as not to restrict the photosynthetic process. The top growth of the fodder x sugarbeet and fodder beet types was similar to that of the sugarbeets throughout the season so it can be concluded that the water requirements of various genotype types would also be similar. However, the increased top growth early in the season and the late season loss in active leaf area by the fodder beet types, may vary slightly their evapotranspiration rates during that part of the season as compared with sugarbeet types.

There were no N-by-water or genotype-by-water interactions in this experiment. All of the genotypes were affected similarly by water stress at each of the N levels (Table 7). The major effect on each genotype-by-water stress was to reduce the root yield by an average 7%, maintain the sucrose, total sugars, and dry matter concentrations at the  $M_1$  irrigation level, and reduce root and total dry matter yields by an average 5%. Consequently, the total sugar and potential alcohol yields were reduced by water stress an average 7%. This reduction in total sugar yield compares well with the 2% sucrose yield reductions in 1977 (3), 6% in 1978 (3), and 5% in 1979 (4) caused by the irrigation cutoff on 1 August. However, it becomes apparent from comparing sugarbeet yields and yield reductions between

Beta vulgaris		Pettole $\text{NO}_3\text{-N}$	Pettole $\text{NO}_3\text{-N}$ (21 August) <sup>†</sup>	Beta vulgaris	Pettole $\text{NO}_3\text{-N}$ (21 August) <sup>†</sup>	Genotype	$\text{kg N/ha}$						
GMD2	110	2180	196	Monrosa	100	1680	ppm	ppm	ppm	ppm	ppm	ppm	ppm
AH10	210 <sup>‡</sup>	2070	196	Monoblanca	110	1360	ppm	ppm	ppm	ppm	ppm	ppm	ppm
LHY-1	280	1960	196	Pajibjerg Kortsro	250	2360	ppm	ppm	ppm	ppm	ppm	ppm	ppm
LHS-1	360	2100	196	Roca	110	1570	ppm	ppm	ppm	ppm	ppm	ppm	ppm

<sup>†</sup>  $M_1$  irrigation level. <sup>‡</sup> Check plots=100 ppm.

GMD2  
AH10  
LHY-1  
LHS-1

110  
210<sup>‡</sup>  
280  
360

2070  
1960  
196  
196

years caused by water stress, that yield reductions increase as root and sucrose yields increase. Significant yield reductions caused by water stress only occurred during years or on fields where the root and sucrose yields were higher than average.

Water use by the sugarbeet fodder x sugarbeet, and fodder beet types was quite similar under each irrigation treatment (Figure 2A). When adequate soil water was present, evapotranspiration (ET), estimated from water depletion of the profile

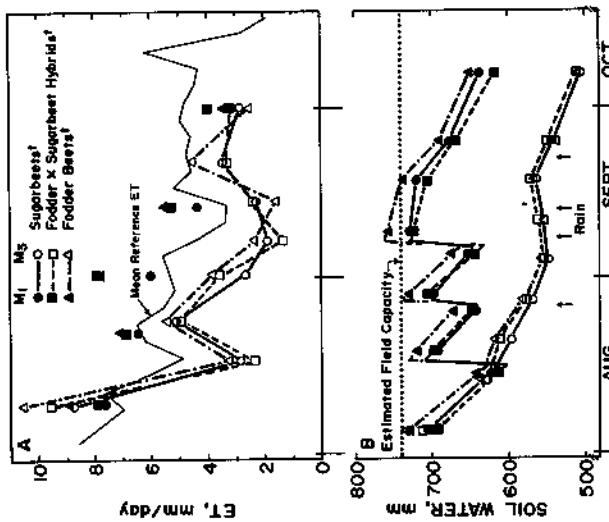


Figure 2. Measured soil water content and evapotranspiration ET, and estimated field capacity for 1980 [Estimated field capacity determined at 0.33 bar. Mean (3-day) reference ET (alfalfa) determined by methods of Wright (23)].  
=GWD2, AH10, LHY-1, and LHS-1; Fodder x Sugarbeet hybrids = Monorosa and Monoblanc; Fodder beets = Pajibjergr Korsroe and Rota.

using average neutron probe measurements for the upper and lower parts of the field on each genotype, followed a rather consistent pattern and was similar to those found in 1977 and 1978 (3), and 1979 (4), as compared with the potential or reference ET (alfalfa, *Medicago sativa* L.) as determined by J. L. Wright using described procedures (23). Evapotranspiration generally decreased after water cutoff on all genotypes as the soil water was depleted and as the potential ET decreased because of lower solar radiation and air temperatures. Evapotranspiration increased after significant effective rainfall on the water cutoff treatment because of the increased surface water. Compared with

Genotype	Root Yield			Total Sugars			Alcohol			Root DM			DM Yd.			Total DM		
	M <sub>1</sub>	M <sub>3</sub>	M <sub>1</sub>	M <sub>1</sub>	M <sub>3</sub>	M <sub>1</sub>	M <sub>1</sub>	M <sub>3</sub>										
GWD2	86.0	* 78.7	17.9	17.5	15.4	* 13.8	9.16	* 8.22	23.7	23.4	27.0	* 24.2	—	—	—	—	—	
AH10	79.8	* 74.8	17.9	17.6	14.0	* 13.1	8.37	* 7.81	24.1	24.0	26.3	* 24.4	—	—	—	—	—	
LHY-1	87.0	* 82.3	18.1	17.9	15.8	* 14.7	9.40	* 8.76	24.1	24.0	26.9	* 25.3	—	—	—	—	—	
LHS-1	73.0	69.1	19.9	19.4	14.5	* 13.4	8.62	* 7.98	26.1	26.8	25.0	24.4	—	—	—	—	—	
Monorosa	98.5	* 90.8	14.7	15.0	14.4	13.6	8.58	8.08	19.7	20.1	24.7	* 23.3	—	—	—	—	—	
Monoblanc	112.0	* 100.8	13.7	13.6	15.3	* 13.7	9.14	* 8.18	18.3	18.7	25.7	* 23.8	—	—	—	—	—	
Pajibjergr Korsroe	131.8	* 124.5	11.6	11.6	15.2	14.5	9.06	8.63	15.9	16.4	24.9	24.4	—	—	—	—	—	
Rota	130.2	* 124.4	10.1	10.1	12.8	12.5	12.5	7.63	13.7	14.1	21.3	21.0	—	—	—	—	—	
LSD (0.05)	4.7	4.7	0.5	0.5	0.5	0.5	0.9	0.53	0.53	0.5	1.3	1.3	—	—	—	—	—	

\* Significant interaction at p = 0.05; between M Leve<sub>1</sub> means = significant difference means at p = 0.05.

Genotype	Root Yield			Total Sugars			Alcohol			Root DM			DM Yd.			Total DM		
	M <sub>1</sub>	M <sub>3</sub>	M <sub>1</sub>	M <sub>1</sub>	M <sub>3</sub>	M <sub>1</sub>	M <sub>1</sub>	M <sub>3</sub>										
GWD2	86.0	* 78.7	17.9	17.5	15.4	* 13.8	9.16	* 8.22	23.7	23.4	27.0	* 24.2	—	—	—	—	—	
AH10	79.8	* 74.8	17.9	17.6	14.0	* 13.1	8.37	* 7.81	24.1	24.0	26.3	* 24.4	—	—	—	—	—	
LHY-1	87.0	* 82.3	18.1	17.9	15.8	* 14.7	9.40	* 8.76	24.1	24.0	26.9	* 25.3	—	—	—	—	—	
LHS-1	73.0	69.1	19.9	19.4	14.5	* 13.4	8.62	* 7.98	26.1	26.8	25.0	24.4	—	—	—	—	—	
Monorosa	98.5	* 90.8	14.7	15.0	14.4	13.6	8.58	8.08	19.7	20.1	24.7	* 23.3	—	—	—	—	—	
Monoblanc	112.0	* 100.8	13.7	13.6	15.3	* 13.7	9.14	* 8.18	18.3	18.7	25.7	* 23.8	—	—	—	—	—	
Pajibjergr Korsroe	131.8	* 124.5	11.6	11.6	15.2	14.5	9.06	8.63	15.9	16.4	24.9	24.4	—	—	—	—	—	
Rota	130.2	* 124.4	10.1	10.1	12.8	12.5	12.5	7.63	13.7	14.1	21.3	21.0	—	—	—	—	—	
LSD (0.05)	4.7	4.7	0.5	0.5	0.5	0.5	0.9	0.53	0.53	0.5	1.3	1.3	—	—	—	—	—	

Yield<sub>1</sub>s; DM = dry matter, Yd = yield, mc = metric tons, kL = kiloliters. Total sugars (% and yield), percent alcohol yield, root percent dry matter, and total plant dry matter for root yield, sucrose yield, and alcohol yield.

Table 7. Interaction means for each genotype (M<sub>1</sub>) level and moisture level for root yield, sucrose yield, and alcohol yield.

the normal irrigation treatment, the water stress treatment reduced total ET by 134 mm when all genotypes were averaged.

The available water in the profile on the water cutoff treatment decreased steadily on all sugarbeet and fodder beet types during August and September without either irrigation or significant rainfall (Figure 2B). The water used for ET came mainly from above the hard layer when irrigation water was adequate. However, as the surface soil dried and approached the wilting range on the irrigation water cutoff treatment as much as 70 to 80% of the water used for ET came from the hard layer and below (60 to 230 cm). The total available water in these silt loam soils between the estimated field capacity (0.33 bar) and the maximum extraction (about 10 bar) is about 260 mm (3). The average total water used by the genotypes between the estimated field capacity and maximum extraction in this experiment was 230 mm of water on the irrigation water cutoff treatment. This demonstrated that 230 mm of available water was present in this soil and that an additional 30 mm of water may become available. Past research with sugarbeets has shown that very little reduction in sucrose yield can be expected by 1 August water cutoff if soil profile is filled with water about 1 August on soils where the usable soil water reservoir is at least 200 mm.

This experiment showed that the different genotypes act similarly under mid- to late-season moisture stress. All genotypes were able to use effectively the soil water below the hard layer probably by root penetration of the hard layer through small cracks or holes made by roots from previous crops having a stronger rooting system such as alfalfa. Mid- to late-season water stress reduced the irrigation water use by all genotypes an average of 24% as compared with the normally irrigated areas with very little total sugar yield reduction. The drought tolerance of the genotypes would allow their growth for alcohol production on land areas of marginal value for most food crops due to limited water for irrigation.

The results of this experiment showed clearly that the higher root yielding fodder beet types had no fermentable sugar yield or other advantages over the higher yielding commercial

or experimental sugarbeet types for alcohol production. In addition, fodder beet types have many disadvantages such as: 1) lower sugar concentration in the root that is due mainly to increased water content; 2) increased weight and volume per unit of sugar that increases the cost of harvesting, hauling to the factory, storage in piles, and processing; 3) increased moisture content decreases the time the roots can be stored in piles; 4) difficulty in defoliation and harvesting using present day equipment because a major part of the storage root is above the soil surface; 5) lack of resistance to curly top; 6) the multigerm seed would prevent mechanical thinning and add to the cost of proper plant spacing.

Using alcohol as a renewable energy source will depend upon the cost in relation to other renewable and nonrenewable energy sources and its priority in relation to food products for land use for biomass production. If alcohol is used to supply part of U. S. energy needs, using the higher yielding sugarbeet varieties presently grown or those developed in breeding programs, could make this crop a primary feedstock source for alcohol production.

#### SUMMARY

Several genetically diverse *Beta vulgaris* genotypes were grown in a field experiment on a Portneuf silt loam soil to evaluate their fermentable sugar yield potential under moderate and high N levels, and normal and mid- to late-season soil water deficit and plant water stress. The genotypes varied widely in their root yield, sucrose and total sugar concentrations with both N and irrigation water levels. There was an inverse linear relationship between root yield and total sugar concentration, resulting in similar total sugar yields and potential alcohol production among most genotypes. The N requirements for maximum sugar yields were slightly higher for fodder beets as compared with sugarbeets. This resulted from the extra dry matter production with increased N level being mainly partitioned to the sugarbeet tops; whereas with fodder beet types, the extra dry matter and growth were equally divided between the tops and roots. All genotypes were similarly affected by mid- to

late-season water stress and showed an average 7% total sugar yield reduction. The results showed that despite the higher root yields, fodder beet types had no sugar yield or other advantage over the better sugarbeets. Current high yielding sugarbeet hybrids are recommended as feedstock source for alcohol production.

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#### LITERATURE CITED

- (1) Bremner, J. M. 1965. Inorganic forms of nitrogen. In C. A. Black (ed.) *Methods of Soil Analyses*. Part 2. Agron. 9:1179 - 1237. Am. Soc. of Agron., Madison, Wisconsin.
- (2) Carter, J. N., M. E. Jensen, and S. M. Bosma. 1974. Determining nitrogen fertilizer needs for sugarbeets from residual soil nitrate and mineralizable nitrogen. Agron. J. 66:319 - 323.
- (3) Carter, J. N., M. E. Jensen, and D. J. Traveller. 1980. Effect of mid- to late-season water stress on sugarbeet growth and yield. Agron. J. 72:806 - 815.
- (4) Carter, J. N., D. J. Traveller, and R. C. Rosenau. 1980. Root and sucrose yields of sugarbeets as affected by mid- to late-season water stress. J. Am. Soc. Sugar Beet Technol. 20:583 - 596.
- (5) Carter, J. N., D. T. Westermann, and M. E. Jensen. 1976. Sugarbeet yield and quality as affected by nitrogen level. Agron. J. 68:49 - 55.
- (6) Doney, D. L. and J. C. Theurer. 1980. Alcohol fuel from sugarbeet. Utah Sci. 41:40 - 43.
- (7) Doney, D. L. 1980. Sugarbeet - fodderbeet as a source of alcohol fuel. Proc. of Bio-Energy '80, Atlanta, Georgia. 21-24 April 1980, pp. 95 - 97.
- (8) Dunn, J. S., D. F. Henderson, D. J. Painter, P. E. Stock, and R. J. Young. 1978. Ethanol produced from fodder beet. New Zealand Agr. Eng. Inst., Lincoln College, Canterbury, NZ.
- (9) Giles, J. F., J. R. Reuss, and A. E. Ludwick. 1975. Prediction of nitrogen status of sugarbeets by soil analysis. Agron. J. 67:454 - 459.
- (10) Hills, F. J. and A. Ulrich. 1976. Soil nitrate and the response of sugarbeets to fertilizer nitrogen. J. Am. Soc. Sugar Beet Technol. 19:118-124.
- (11) James, D. W., A. W. Richards, W. H. Weaver, and R. L. Reeder. 1971. Residual soil nitrate measurements as a basis for managing nitrogen fertilizer practices for sugarbeets. J. Am. Soc. Sugar Beet Technol. 16: 313 - 322.
- (12) Jensen, M. E., and L. J. Erie. 1971. Irrigation and water management. pp. 189 - 222. In R. T. Johnson, J. T. Alexander, G. E. Rush, and G. R. Hawkes (eds.) *Advances in Sugarbeet Production: Principles and Practices*. The Iowa State Univ. Press Ames, Iowa.
- (13) Jensen, M. E., D. C. Robb, and C. E. Franzoy. 1970. Scheduling irrigations using climatic-crop-soil data. J. Irrig. Drain. Div., Am. Soc. Civ. Eng. 96(IRI):25-38.
- (14) Kelley, J. D., and A. Ulrich. 1966. Distribution of nitrate nitrogen in the blades and petioles of sugarbeets grown at deficient and sufficient levels of nitrogen. J. Am. Soc. Sugar Beet Technol. 14:106 - 116.
- (15) Loomis, R. S. 1979. Ideotype concepts for sugarbeet improvement. J. Am. Soc. Sugar Beet Technol. 20:323-342.
- (16) McGinnis, R. A. 1971. Analysis of sugar content. p. 78 In R. A. McGinnis (ed.) *Beet-Sugar Technology*. Beet Sugar Dev. Found., Ft. Collins, Colo.
- (17) Milham, P. J., A. S. Awad, R. E. Paull, and J. H. Bull. 1970. Analysis of plants, soils, and water for nitrate by using an ion-selective electrode. Analysis 95:751 - 757.
- (18) Miller, D. E., and J. S. Aarsstad. 1976. Yields and sugar content of sugarbeets as affected by deficit high-frequency irrigation. Agron. J. 68:231 - 234.
- (19) Surface, G. T. 1910. *The Story of Sugar*. P. Appleton and Co., New York, N.Y.
- (20) Ulrich, A. 1950. Critical levels of sugar beets estimated from analysis of petioles and blades, with special reference to yields and sucrose concentrations. Soil Sci. 69:291 - 309.
- (21) Westermann, D. T., G. E. Leggett, and J. N. Carter. 1977. Phosphorus fertilization of sugarbeets. J. Am. Soc. Sugar Beet Technol. 19:262 - 269.
- (22) Winter, S. R. 1980. Suitability of sugarbeets for limited irrigation in a semi-arid climate. Agron. J. 72:118-123.
- (23) Wright, J. L. 1982. New evapotranspiration crop coefficients. J. Irrig. Drain. Div., Am. Soc. Civ. Eng. 108(IRR2):57 - 74.